

Diastereoisomeric ichthyotoxic acylglycerols from the dorsum of two geographically distinct populations of *Archidoris nudibranchs*

E. Zubía¹, M. Gavagnin, A. Crispino, E. Martínez^a, J. Ortea^a and G. Cimino

Istituto per la Chimica di Molecole di Interesse Biologico del C.N.R., Via Toiano 6, I-80072 Arco Felice, Napoli (Italy), and ^aDepartamento de Biología de Organismos y Sistemas, Universidad de Oviedo, C/J. Arias de Velasco, Oviedo (Spain)

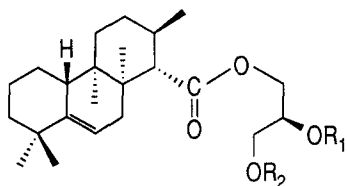
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Abstract. Two marine opisthobranchs, *Archidoris tuberculata* from Asturias (N. Spain) and *Archidoris carvi* from Patagonia (S. Argentina), contain in their dorsum diastereoisomeric ichthyotoxic acylglycerols esterified in position 1-*sn* by antipodal diterpenoid acids.

Key words. Acylglycerols; ichthyotoxins; nudibranchs; marine molluscs.

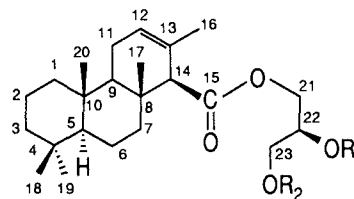
Chemical studies of the defensive strategies of the apparently unprotected marine nudibranchs have discovered many ecologically relevant molecules^{2,3}, which not only play a part in defensive strategies but often possess other basic biological functions⁴. Among recent exam-

ples are studies of the multifunctional drimane sesquiterpenoids from Dendrodorididae species⁵, of prostaglandin lactones from *Tethys fimbria*⁶ and, finally, of some diacylglycerols, verrucosins (**1**, **2**), from *Doris verrucosa*⁷. The interesting biological properties of verru-



1 R₁ = H, R₂ = COCH₃

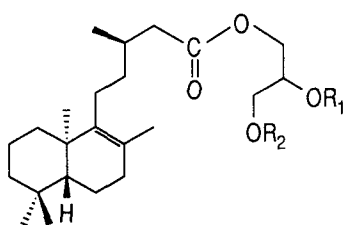
2 R₁ = COCH₃, R₂ = H



3 R₁ = H, R₂ = COCH₃

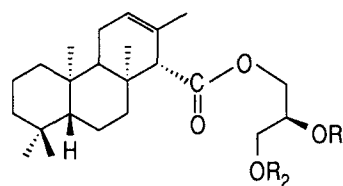
4 R₁ = COCH₃, R₂ = H

5 R₁ = R₂ = H



6 R₁ = H, R₂ = COCH₃

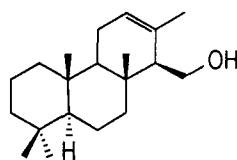
7 R₁ = COCH₃, R₂ = H



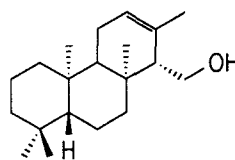
8 R₁ = H, R₂ = COCH₃

9 R₁ = COCH₃, R₂ = H

10 R₁ = R₂ = H



11



12

cosins (**1**, **2**), which are highly ichthyotoxic to *Gambusia affinis*^{8,9} and are activators of protein kinase C¹⁰, could be due to their unusual structure^{7,11}, featuring a rearranged isocopalane¹² diterpenoid ester in position C-1 of *sn*-glycerol. Structurally related acyl glycerols were described previously from British Columbian *Archidoris montereyensis*^{13,14} and *Archidoris odhneri* (**3–5**)¹⁴, and more recently from Antarctic *Austrodoris kerguelensis* (**6, 7**)¹⁵. Even though biosynthetic experiments were successful only with *A. montereyensis* and *A. odhneri*, it seems an attractive hypothesis that there is a small group of Eudoridoidea nudibranchs, belonging to closely related families, which is able to produce potentially defensive allomones by de novo biosynthesis of acylglycerols.

In this paper we report studies on two further Eudoridoidea nudibranchs, *Archidoris tuberculata* (Müller, 1778) and *Archidoris carvi* (Marcus, 1955). The former possesses the same metabolites (**3–5**) as *A. montereyensis*, whereas *A. carvi* contains two new glycerides (**8, 9**) characterized by a diterpenoid residue enantiomeric with that of **3–5**.

Specimens of *A. tuberculata* (16 individuals) were collected at Rio del Eo (Asturias, North Spain) in the intertidal zone from December, 1990, to April, 1991. Samples were immediately frozen and stored at low temperature until February, 1992. Mantles and viscera were separately extracted with acetone. The TLC (SiO₂, *n*-hexane/diethyl ether, 1:1) chromatographic

pattern of the diethyl ether-soluble fraction of the acetone extract of the mantles was dominated by two compounds (*R_f* 0.35 and 0.30) which were completely absent in the extract of the viscera. Subsequent separation on a preparative column (SiO₂, petroleum ether/diethyl ether gradient) and by HPLC (μ -Porasil, *n*-hexane/ethyl acetate, 85:15) led to two compounds (5.5 mg and 2.5 mg) identical to **3** and **4**, already described¹⁴. Superior NMR facilities which became available later allowed more complete assignments (table) which agreed well for the diterpenoid part with the NMR values previously assigned¹⁶. Chromatography also led to the isolation of a more polar compound identical with **5**, and of a less polar acylglycerol (*R_f* 0.6, TLC), probably esterified by a bicyclic diterpenoid acid residue, whose structure remains to be determined. Specimens of *A. carvi*¹⁷ (7 individuals) were collected at Punta Pardelas (Peninsula of Valdes, South Argentina) by snorkeling, at a depth of 1–10 m during November, 1991. The molluscs were stored, dissected and analyzed analogously to *A. tuberculata*. The ether-soluble fraction (80 mg) from the acetone extract of the mantles displayed a TLC chromatographic pattern almost identical with that of the corresponding extract from *A. tuberculata*. After two chromatographic steps [(1) SiO₂ column, gradient of *n*-hexane/diethyl ether mixtures; (2) μ -Porasil, *n*-hexane/diethyl ether, 85:15] two diacylglycerols **8** (2.2 mg) and **9** (0.8 mg) were obtained.

Table. ¹H- and ¹³C-NMR data of **3**, **4**, **8** and **9**^a

C	$\delta^1\text{H}^b$				$\delta^{13}\text{C}(\text{m})^b$	
	3	4	8	9	3	8
1	0.80 1.61	0.80 1.61	0.80 1.62	0.80 1.62	39.84(t)	39.85(t)
2	1.38 1.58	1.38 1.58	1.38 1.57	1.38 1.57	18.60*(t)	18.61*(t)
3	1.14 1.38	1.14 1.38	1.14 1.38	1.13 1.38	41.82(t)	41.83(t)
4	-	-	-	-	33.14(s)	33.0(s)
5	0.84	0.84	0.85	0.85	56.41(d)	56.42(d)
6	1.38 1.58	1.38 1.58	1.38 1.57	1.38 1.57	18.44*(t)	18.45*(t)
7	1.38 1.70	1.38 1.68	1.38 1.70	1.38 1.68	41.82(t)	41.83(t)
8	-	-	-	-	37.39(s)	37.5(s)
9	1.16	1.15	1.16	1.15	54.24(d)	54.24(d)
10	-	-	-	-	36.58(s)	36.5(s)
11	1.95	1.95	1.95	1.95	22.64(t)	22.65(t)
12	5.53	5.53	5.53	5.52	124.32(d)	124.36(d)
13	-	-	-	-	128.45(s)	128.46(s)
14	2.96	2.95	2.96	2.94	62.51(d)	62.51(d)
15	-	-	-	-	173.02(s)	§
16	1.60	1.60	1.60	1.60	21.16(q)	21.19(q)
17	0.94*	0.93*	0.95*	0.93*	15.57(q)	15.56(q)
18	0.81	0.83	0.81	0.81	21.64(q)	21.64(q)
19	0.86	0.86	0.86	0.86	33.40(q)	33.41(q)
20	0.90*	0.90*	0.90*	0.90*	15.72(q)	15.72(q)
21	4.12 4.22 ^c	4.27 4.34	4.12–4.24 ^c	4.23–4.35	64.83(t)	64.92(t)
22	4.10	5.07	4.12	5.10	68.37(d)	68.40(d)
23	4.12 4.22 ^c	3.76	4.12–4.24 ^c	3.76	65.30(t)	65.29(t)
OCOMe	2.11	2.11	2.11	2.10	20.77(q)	20.78(q)

^aBruker AMX-500 spectrometer; CDCl₃; chemical shifts referred to CHCl₃ at 7.26 ppm and to C₂DCl₃ at 77.00 ppm.

^bThe assignments were aided by ¹H-¹H COSY and ¹H-¹³C HETCOR. ^cUnresolved 4 proton multiplets. §Not detected.

*Starred values in the same column can be interchanged.

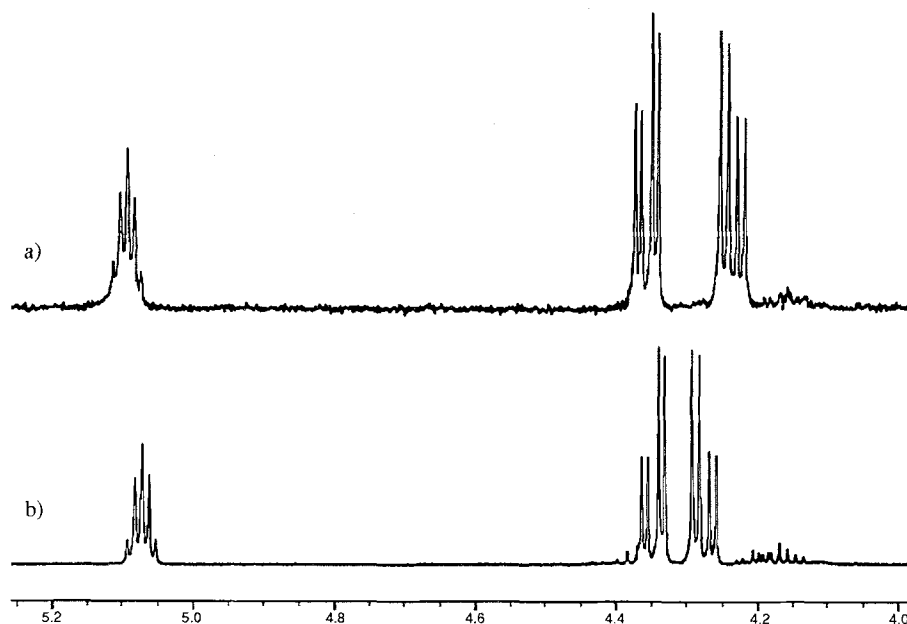


Figure 1. Partial ^1H -NMR of **9**(a) and **4**(b).

The spectral data of **8** and **9**¹⁸ were almost identical to those of **3** and **4**. However, detailed careful comparison of the ^1H -NMR spectra of **4** and **9** revealed diagnostic differences (fig. 1) in the chemical shifts of the H-21 and H-22.

Similarly, **3** and **8** exhibited characteristic ^1H -NMR profiles which were different, but too complex to analyze as all glycerol protons resonate between δ 4.10 and 4.24. Fortunately, slight shifts were observed in their ^{13}C -NMR spectra for C-21 (δ 64.83 for **3** and δ 64.92 for **8**) (table). These data supported diastereoisomeric relationships between the two pairs **3–8** and **4–9**, which could be due to opposite absolute stereochemistry, either of the diterpenoid acid or of the glycerol moiety. While the optical rotations of **3** and **4** are negative, as already reported¹⁴, the corresponding values for **8**, $[\alpha]_{\text{D}} + 21.9^\circ$ ($c = 0.22$, CHCl_3), and **9**, $[\alpha]_{\text{D}} + 66.9^\circ$ ($c = 0.07$, CHCl_3), were both positive, thereby suggesting opposite stereochemistry of the diterpenoid residue. The CD spectra (fig. 2) displayed at the same wavelength (~ 214 nm) negative curves for **3**, **4** and **5**, and positive for **8**, **9** and their deacetyl derivative **10**. Oxidation of **3** and **8** with Jones' reagent yielded ketones displaying identical ^1H -NMR [0.82, 0.86, 0.91, 0.95, 2.18 (s, 3H each); 3.04 (bs, H-14); 4.75 (m, H-21 and H-23), 5.55 (m, H-12)] spectra but coherent opposite CD profiles. Finally, reduction of **3** and **8** with LiAlH_4 yielded alcohol **11**^{13,19}, $[\alpha]_{\text{D}} - 12^\circ$ ($c = 0.25$, CHCl_3); lit. $[\alpha]_{\text{D}} - 9^\circ$, and its enantiomer **12**, $[\alpha]_{\text{D}} + 10^\circ$ ($c = 0.04$, CHCl_3), thus confirming the suggested isocopalane skeleton of the diterpenoid acid residue of **8**²⁰.

It is worth noting that probably only **4** and **9** are natural products, whereas **3** and **8** might arise from an

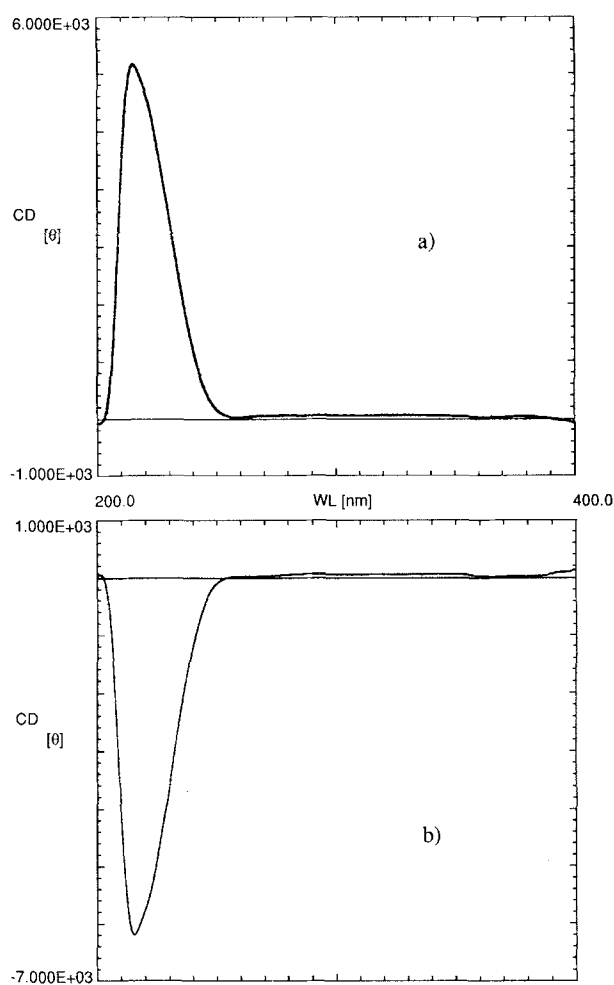


Figure 2. CD spectra of **8**(a), $[\theta]_{214} = 5412$, and **3**(b), $[\theta]_{214} = -6195$.

easy *trans*-acetylation. However, bearing in mind the stereospecific numbering for the derivatives of glycerols, both **4** and **9** are 1,2-diacyl-*sn*-glycerols. Enantiomeric terpenoids are rather common among sesquiterpenoids²¹, relatively rare among diterpenoids²², and extremely rare among the superior terpenoids where there are also some inexact reports, e.g. *ent*-12-*epi*-deoxoscalarin²³.

In conclusion, it seems that *Archidoris*, *Doris* and *Austrodoris* nudibranchs have elaborated a defensive strategy which utilizes acylglycerols as defensive allomones. Preliminary bioassays have revealed strong ichthyotoxicity of **3** and **4** at 10 and 1 ppm against *Gambusia affinis*.

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18 Spectral data for **8**: IR (film) 3450, 1738 cm⁻¹; ¹H-NMR and ¹³C-NMR in table. MS, *m/z* 420(0.5), 402(2), 347(2), 286(100), 192(67), 177(67), 117(47). Spectral data for **9**: IR (film) 3400, 1738 cm⁻¹; ¹H-NMR in table. MS, *m/z* 420(1), 402(2), 347(2), 286(100), 192(61), 177(71), 117(70).

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20 Surprisingly, recording CD spectra of verrucosin-A and -B (**1** and **2**) positive curves, analogous with those of **8** and **9**, were obtained. An erroneous observation of the X-ray model of **2** led to assign¹¹ a rearranged *ent*-isocopalane skeleton to the diterpenoid residue whereas, correctly, verrucosins should derive from an isocopalane precursor as shown by formulas **1** and **2**.

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